Cytomegalovirus (CMV) IgG

Catalog No: DT50014 96 Tests

NAME AND INTENDED USE
The D-TEK CMV IgG ELISA is intended for use in the evaluation of patients with suspected CMV infection.

SUMMARY AND EXPLANATION OF THE TEST
Cytomegalovirus (CMV) is a member of the herpes group of viruses. Most adults and children who catch CMV have no symptoms and are not harmed by the virus. CMV infection is of clinical significance primarily in pregnant women, newborn infants with possible congenital infection, immunosuppressed transplant patients and individuals with AIDS. CMV is so prevalent as over 60% of people catch the infection at some time in their lives.

Significant increases in CMV IgG antibody by ELISA suggest recent infection or reactivation of a latent CMV infection.

ELISA can detect CMV IgM antibody in both primary CMV infections (93-100%) and in reactivated infection (40%). An IgM response may be reduced or absent with active infection. In transplant patients the CMV infection can be associated with higher morbidity and mortality.

PRINCIPLE OF THE TEST
Diluted patient serum is added to wells coated with purified antigen. IgG specific antibody, if present, binds to the antigen. All unbound materials are washed away and the enzyme conjugate is added to bind to the antibody-antigen complex, if present. Excess enzyme conjugate is washed off. The plate is incubated to allow the hydrolysis of the substrate by the enzyme. The intensity of the color generated is proportional to the amount of IgG specific antibody in the sample.

MATERIALS PROVIDED
1. Microwell Strips: CMV antigen coated wells (12 x 8 x 1 wells)
2. Sample Diluent: 1 bottle (22 mL). Ready to use.
3. Calibrator: Yellow Cap. (1.50 mL/vial). Ready to use.
4. Positive Control: Red Cap. (1.50 mL/vial). Ready to use.
5. Negative Control: Blue Cap. (1.50 mL/vial). Ready to use.
6. Enzyme Conjugate: 1 bottle (12 mL). Ready to use.
7. TMB Substrate: 1 bottle (12 mL). Ready to use.
8. Stop Solution: 1N H2SO4; 1 bottle (12 mL). Ready to use.
9. Wash Concentrate: 1 bottle (50 mL), 20X concentrate.

STORAGE AND STABILITY
1. Store the kit at 2-8 °C.
2. Keep microwells sealed in a dry bag with desiccants.
3. The reagents are stable until expiration of the kit.
4. Do not expose test reagents to heat, sun or strong light during storage or usage.

WARNINGS AND PRECAUTIONS
1. Potential biohazardous materials: The calibrator and controls contain human source components which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, there is no test method that can offer complete assurance that HIV, Hepatitis B virus or other infectious agents are absent. These reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, "Biosafety in Microbiological and Biomedical Laboratories." 1984.
2. Optimal results will be obtained by strict adherence to the test protocol. Precise pipetting as well as following the exact time and temperature requirements is essential.
3. Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.
4. The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed.
5. Control sera and sample diluent contain preserved with sodium azide. Sodium azide may react with lead and copper plumbing to form explosive metal azide. On disposal, flush with a large volume of water.

SPECIMEN COLLECTION AND HANDLING
1. Collect blood specimens and separate the serum.
2. Specimens may be refrigerated at 2–8 °C for up to seven days or frozen for up to six months. Avoid repetitive freezing and thawing of samples.

PREPARATION FOR ASSAY
1. Bring all specimens and kit reagents to room temperature (20-25 °C) and gently mix.
2. Prepare washing buffer by adding the contents of the bottle (50 mL, 20X Wash concentrate) to 950 mL of distilled or deionized water in one-liter container. Store at room temperature.

ASSAY PROCEDURE
1. Place the desired number of coated strips into the holder.
2. Negative control, positive control, and calibrator are ready to use.
3. Prepare 1:21 dilution of test samples, by adding 10 µL of the sample to 200 µL of sample diluent. Mix well.
4. Dispense 100 µL of diluted sera, calibrator and controls into the appropriate wells. For the reagent blank, dispense 100 µL sample diluent in 1A well position. Tap the holder to remove air bubbles from the liquid and mix well. Incubate for 20 minutes at room temperature.
5. Remove liquid from all wells. Repeat washing three times with wash buffer.
6. Dispense 100 µL of enzyme conjugate to each well and incubate for 20 minutes at room temperature.
7. Remove enzyme conjugate from all wells. Repeat washing three times with wash buffer.
8. Dispense 100 µL of TMB substrate solution and incubate for 10 minutes at room temperature.
9. Add 100 µL of 1N H2SO4 to stop reaction.
10. Read O.D. within 30 min at 450 nm using microwell reader.
CALCULATION OF RESULTS
1. Check Calibrator Factor (CF) value on the calibrator bottle. This value might vary from lot to lot. Make sure you check the value on every kit.
2. Calculate the cut-off value: Calibrator OD x Calibrator Factor (CF).
3. Calculate the Ab (Antibody) Index of each determination by dividing the O.D. value of each sample by cut-off value.

Example of typical results:
Calibrator mean OD = 0.8
Calibrator Factor (CF) = 0.5
Cut-off Value = 0.8 x 0.5= 0.400
Positive control O.D. = 1.2
Ab Index = 1.2 / 0.4 = 3
Patient sample O.D. = 1.6
Ab Index = 1.6 / 0.4 = 4.0

QUALITY CONTROL
The test run may be considered valid provided the following criteria are met:
1. The O.D. of the Calibrator should be greater than 0.250.
2. The Ab index for Negative control should be less than 0.9.
3. The Ab Index for Positive control should be greater than 1.2.

INTERPRETATION
The following is intended as a guide to interpretation of CMV IgG test results; each laboratory is encouraged to establish its own criteria for test interpretation based on sample populations encountered.

1. Antibody Index Interpretation
   - <0.9 No detectable antibody to CMV IgG by ELISA.
   - 0.9-1.1 Borderline positive. Follow-up testing is recommended if clinically indicated.
   - >1.1 Indicative of current or previous CMV infection.

2. Converting of Ab Index to IU/mL
   As an option, Ab index may be converted to IU/mL by multiplying Ab index by 1. IU/mL values may then be interpreted as follows:
   - <0.9 IU/mL No detectable IgG antibody to CMV

LIMITATIONS OF THE TEST
1. The test results obtained using this kit serve only as an aid to diagnosis and should be interpreted in relation to the patient's history, physical findings and other diagnostic procedures.
2. Lipemic or hemolyzed samples may cause erroneous results.

PERFORMANCE CHARACTERISTICS
1. Sensitivity and Specificity
   180 patient sera were tested by this CMV IgG ELISA and a reference ELISA method. 112 sera were positive and 61 were negative by both methods (96% agreement). The results are summarized below:

<table>
<thead>
<tr>
<th></th>
<th>Reference ELISA Kit</th>
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</thead>
<tbody>
<tr>
<td>Serum</td>
<td>+</td>
</tr>
<tr>
<td>CMV IgG ELISA</td>
<td></td>
</tr>
<tr>
<td>Reference</td>
<td>112</td>
</tr>
<tr>
<td>Intra-Assay</td>
<td>3</td>
</tr>
<tr>
<td>Inter-Assay</td>
<td>115</td>
</tr>
</tbody>
</table>

2. Precision
   Intra-Assay Study
<table>
<thead>
<tr>
<th>Serum</th>
<th>No. of Replicates</th>
<th>Mean</th>
<th>S.D.</th>
<th>C.V. %</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>16</td>
<td>0.97</td>
<td>0.019</td>
<td>1.96</td>
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<tr>
<td>2</td>
<td>16</td>
<td>0.64</td>
<td>0.025</td>
<td>3.90</td>
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<tr>
<td>3</td>
<td>16</td>
<td>0.09</td>
<td>0.007</td>
<td>7.77</td>
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   Inter-Assay Study
<table>
<thead>
<tr>
<th>Serum</th>
<th>No. of Replicates</th>
<th>Mean</th>
<th>S.D.</th>
<th>C.V. %</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>10</td>
<td>1.39</td>
<td>0.14</td>
<td>10.07</td>
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<tr>
<td>2</td>
<td>10</td>
<td>0.6</td>
<td>0.049</td>
<td>8.16</td>
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<tr>
<td>3</td>
<td>10</td>
<td>0.19</td>
<td>0.023</td>
<td>12.10</td>
</tr>
</tbody>
</table>

REFERENCES
2. Gupta CK; Leszczynski J; Gupta RK; Siber GR. An enzyme immunoassay based micro-
5. Schmidt CA; Oettel H; Peng R; Neuhaus P; Blumhardt G; Lohmann R; Willborn F; Ostoff K; Oertel J; Timm H; et al. Comparison of polymerase chain reaction from plasma and buffy coat with antigen detection and occurrence of immunoglobulin M for the demonstration of cytomegalovirus infection after liver transplantation. Transplantation 1995;59(8):1133-8.